The impact of low alcohol consumption on the liver and inflammatory cytokines in diabetic rats


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A – research concept and design; B – collection and/or assembly of data; C – data analysis and interpretation; D – writing the article; E – critical revision of the article; F – final approval of the article

Abstract

**Background.** Diabetes mellitus (DM) and alcohol consumption is still one of the important research models that simulate variable clinical conditions and metabolic diseases, such as alcoholic liver diseases.

**Objectives.** The aim of this study was to evaluate the long-term cumulative effects of low alcohol consumption on the liver tissue, biochemical assays and some inflammatory cytokines in experimentally-induced DM rats.

**Material and methods.** Ethanol was administered in the drinking water (3% v/v) for 30 days to adult male Sprague-Dawley rats, with or without DM induced by streptozocin injection. Histological and biochemical parameters as well as some inflammatory cytokines – interleukin (IL)-4, IL-6, IL-10, and tumor necrosis factor alpha (TNF-α) – were measured.

**Results.** A significant increase in blood glucose level in the combination group was accompanied by a significant decrease in plasma insulin (p < 0.001 vs controls). Hepatic histopathology of the combination group revealed steatosis and fibrosis in addition to a significant increase in the gamma-glutamyltransferase (γ-GT) and alkaline phosphatase (ALP) levels (p < 0.05 and p < 0.001, respectively). A non-high-density lipoprotein (HDL) lipid profile (total cholesterol (TC), triglycerides (TG) and low-density lipoprotein (LDL)) revealed a significant increase in comparison to controls (p < 0.05), while HDL showed no significant change. The IL-4 and IL-6 levels were significantly higher (p < 0.05), while IL-10 and TNF-α revealed non-significant changes.

**Conclusions.** Depletion of the hyperglycemic response in the case of low alcohol consumption in DM rats was associated with elevated plasma cytokines, especially IL-6 and IL-4, which could be a part of a host defense mechanism to repair the hepatic and pancreatic damage through this inflammatory process. The severe liver damage under insult of low alcohol consumption and DM could serve as inhibitory factors in gluconeogenesis and glycogenolysis, with little or no impact on insulin levels.

**Key words:** alcohol, cytokines, diabetes, liver, rat
**Introduction**

Diabetes mellitus (DM) is a public health problem which has reached epidemic proportions worldwide. Insulin-deficient DM is associated with severe complications. Therefore, inflammatory cytokines provide important signals in the pathophysiology and complications of DM, being markedly elevated, and are thought to contribute to several complications (neuropathy, nephropathy, retinopathy, and periodontal lesions). The relationships between plasma inflammatory markers and an increased risk of clinical DM are inconsistent, just like the relationships between these cytokines and other metabolic risk factors, such as dyslipidemia and hepatotoxicity, although hepatic damage is probably a major contributor to dyslipidemia in insulin-deficient DM.

Recent studies have shown that moderate alcohol intake is associated with changes in the levels of several cytokines, especially pro-inflammatory ones – interleukin (IL)-6 and tumor necrosis factor alpha (TNF-α) – in different physiological and pathological states.

Interleukin-6 protects against alcoholic liver injury via the activation of the signal transducer and activator of transcription 3 (STAT3), regulating liver fibrosis and inflammation, and promoting liver regeneration. While IL-10 inhibits alcoholic liver inflammation via the activation of STAT3 in Kupffer cells/macrophages (targeting immune cells) and the subsequent inhibition of liver inflammation.

Recent studies have suggested that IL-10 may play a dual role in controlling ethanol-induced steatosis and liver injury via the inhibition of pro-inflammatory cytokines, such as TNF-α, thereby decreasing alcoholic liver injury, or via the inhibition of the hepatoprotective cytokine IL-6, thereby potentiating alcoholic liver injury.

Furthermore, it has been suggested that ethanol might modulate the production of these cytokines and their clearance at several sites, including adipose tissue. Nevertheles, data on the relationship between alcohol intake and circulating cytokine levels is scant.

The precise role of inflammatory cytokines in the initiation and progression of pathological processes in DM, and its relationship with low alcohol consumption are not fully clear. However, our study contributed to evaluating the changes of the liver (histologically and biochemically) in addition to evaluating the inflammatory response by measuring proinflammatory (IL-6 and TNF-α) and anti-inflammatory (IL-4 and IL-10) cytokines under insult of low alcohol consumption in streptozotocin (STZ)-induced DM in rats.

**Material and methods**

**Animals**

Twenty-four male Sprague-Dawley rats (8–10 weeks old, weighing 221 ±39 g) were used in the experiments. The rats were purchased from the National Center for Drug Research and Quality Control, Baghdad, Iraq. They were kept in the animal house of the Department of Pharmacology and Toxicology, Pharmacy College, University of Babylon, Al-Hilla, Iraq. They were housed 6 per cage, at a constant room temperature (22 ±1°C) and relative humidity (60–70%), and under a 12-hour light/12-hour dark cycle (light 8:00 am–8:00 pm). Standard food and water (treated or not) were provided ad libitum. All experiments were performed according to the international guidelines of laboratory animal care and the ethical guidelines for investigations on experimental animals (International Society for Applied Ethology).

**Induction of diabetes and alcohol consumption**

The rats were divided into 4 main groups (n = 6 per group) as follows: control (C), alcohol consumption (ACH), diabetic (DM), and alcohol consumption with diabetic (ACH+DM). Diabetes mellitus was induced with a single intraperitoneal injection of STZ (65 mg/kg body weigh (b.w.)), diluted in a 0.1 mol/L citrate solution, pH 4.5 (Sigma-Aldrich), while low alcohol administration was performed ad libitum through drinking water consumption with 3% v/v ethanol (99% pure EtOH) (Sigma-Aldrich, St. Louis, USA) to simulate a dosage ranging from 4.0 to 5.0 g/kg b.w. This dose might be enough to induce peripheral insulin sensitization. The body weight of rats was measured once every 5 days. The rats were sacrificed after 30 days of the experiment.

**Blood sample collection**

On the last day of the experiment, blood was collected from the inferior vena cava and the serum was used to estimate the examined biochemical parameters and inflammatory cytokines.

**Histological examination**

The liver specimens were excised and fixed in 4% formalin, then embedded in paraffin wax as per conventional techniques. The sections cut (4 μm) were stained with hematoxylin and eosin (H&E). The specimens were examined by a double-blinded examiner. The morphological changes produced in the liver were histopathologically estimated under a high power field (×400) microscope.

**Biochemical assays**

In order to assess blood glucose, hepatic functional integrity – serum alanine transaminase (ALT), aspartate transaminase (AST), gamma-glutamyltransferase (γ-GT), and alkaline phosphatase (ALP), as well as lipid profile – total cholesterol (TG), triglycerides (TG), high-density lipoprotein (HDL), and low-density lipoprotein (LDL), a biochemical automatic analyzer was used (Hitachi, Roche, USA).
Plasma insulin and cytokines

Plasma insulin was measured by an enzyme-linked immunosorbent assay (ELISA) kit (EZRMI-13K; Linco Research, St. Charles, USA) and the quantity was recorded in ng/mL. Inflammatory cytokines were also measured by an ELISA kit (R&D Systems, Minneapolis, USA). Interleukin-6 was determined by an R6 000B Quantikine kit, IL-4 by an R4 000 Quantikine kit, IL-10 by an R1 000 Quantikine kit, and TNF-α by an RTA00 kit according to the manufacturer's protocol (R&D Systems); each assay was run with known standards (provided with the kit) to determine the quantity of cytokines in each sample in pg/mL.

Statistical analysis

The data is expressed as means ± standard deviation (SD) and was analyzed using a one-way analysis of variance (ANOVA), followed by multiple comparisons with Bonferroni’s method and Tukey’s honestly significant difference method. The significance level for all analyses was set at a probability p < 0.05. All analyses were performed by GraphPad Prism v. 5.3 for Windows (GraphPad Software, San Diego, USA).

Results

Body weight

The body weight of adult rats was not significantly different in the ACH group compared to the controls, but a marked and significant decrease in weight gain appeared in both the DM and ACH+DM groups between 10 and 30 days of the experimental period (p < 0.05 and p < 0.01 for DM, p < 0.001 for ACH+DM vs controls) (Fig. 1), in addition to a significant increase in the liver weight/body weight ratio in the ACH and ACH+DM groups (p < 0.001 for both groups vs controls) (Fig. 2).

Blood glucose and plasma insulin levels

Blood glucose level was significantly higher in the DM and ACH+DM groups (p < 0.001) in comparison to the controls, while non-significant changes were reported in the ACH group compared to the controls (Fig. 3), in contrast to plasma insulin level, which was significantly lower in both the DM and ACH+DM groups compared to the controls (p < 0.01 and p < 0.001, respectively) (Fig. 4).

Liver enzymes

In the ACH, DM and ACH+DM groups, there is a non-significant increase in the enzymes that used for assessment of liver parenymal inflammation and/or injury (ALT and AST) in comparison with the control with different proportions, while the response is different in the enzymes used for the assessment of cholestatic liver inflammation and/or injury (ALP and γ-GT); the ALP level in the ACH+DM group revealed a statistically significant increase in comparison to all groups (control (p<0.001), ACH and DM (p<0.05)). The γ-GT levels were statistically significantly higher in the ACH+DM group only compared to the controls (p < 0.05) (Table 1).
**Determination of the lipid profile**

The TC and TG levels revealed a significant increase in the ACH+DM group compared to the control group and the ACH group ($p < 0.05$) (Table 2). Low-density lipoprotein level was significantly higher in the ACH+DM group compared to the controls ($p < 0.05$), while in the ACH and DM groups, there was no significant difference compared to the control group. High-density lipoprotein measurement revealed no significant differences in the ACH, DM and ACH+DM groups compared to the control group (Table 2).

**Histopathology**

The histological examination of the ACH group revealed a mild inflammatory reaction with progressive microto macrovesicular steatosis with no fibrosis, while the DM group showed microvesicular steatosis that became marked with some fibrosis (Fig. 5). The livers obtained from rats exposed to STZ and alcohol (ACH+DM group) were enlarged, pale in color, soft, and greasy compared to the livers obtained from rats belonging to the control group. The histological examination of the ACH+DM liver sections revealed micro- and macrovesicular steatosis with degeneration and progressive development of lesions due to the effects of alcohol and STZ. Most liver sections showed an increase in fibrosis with plasma cell infiltrate, causing distortion of the usual concentric arrangement of hepatocytes. Moreover, there was congestion of the portal vessels and sinusoids, and the veins were dilated (Fig. 5).

**Inflammatory cytokines**

The IL-4 and IL-6 levels were significantly higher in the DM and ACH+DM groups compared to the control group ($p < 0.05$ and $p < 0.01$, respectively), while non-significant changes were found in the ACH group in comparison to the controls (Fig. 6A,6B, respectively). Furthermore, the IL-6 level was significantly higher in the DM and ACH+DM groups than in the ACH group ($p < 0.05$ and $p < 0.01$, respectively) (Fig. 6B). Moreover, the IL-10 level was non-significantly higher in all experimental groups than in the control group (Fig. 6C), while the TNF-α level was non-significantly lower in the DM and ACH+DM groups than in the control group (Fig. 6D).

**Discussion**

Diabetes mellitus is still one of the important chronic diseases that have a significant impact on different inflammatory and metabolic markers. Our study focused on some of these markers under insult of low alcohol consumption over 30 days. Liver histological examination in the ACH+DM group after 30 days revealed progressive...
micro- and macrovesicular steatosis with degenerative changes; these hepatic fatty deposits were probably due to the inability of the liver to export triglycerides, which had been previously rationalized by the effects of hypoinsulinemia induced by STZ.

Moreover, mild fibrosis with plasma cell infiltrate and congestion of the portal vessels and sinusoids were found. On the other hand, significant weight loss was observed in the combination group, in spite of the fatty liver, due to progressive and severe liver damage, which affected its function and disturbed the normal metabolism by augmenting the catabolic state. This result is in contrast to the weight gain in the DM-only group that was shown by Chowdhury et al., who detected the changes in weight after the induction by STZ with hyperphagia. Moreover, mild fibrosis with plasma cell infiltrate and congestion of the portal vessels and sinusoids were found. On the other hand, significant weight loss was observed in the combination group, in spite of the fatty liver, due to progressive and severe liver damage, which affected its function and disturbed the normal metabolism by augmenting the catabolic state.

### Table 1. The levels of serum liver enzymes indicative of liver functional integrity in diabetic and non-diabetic rats exposed to low-dose alcohol consumption

<table>
<thead>
<tr>
<th>Experimental group</th>
<th>ALT</th>
<th>AST</th>
<th>ALP</th>
<th>γ-GT</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>351.2 ±26.3</td>
<td>74.3 ±16.1</td>
<td>162.2 ±13.4</td>
<td>0.33 ±0.15</td>
</tr>
<tr>
<td>ACH</td>
<td>373.6 ±37.1</td>
<td>109.8 ±38.6</td>
<td>383.4 ±73.2</td>
<td>1.6 ±0.24</td>
</tr>
<tr>
<td>DM</td>
<td>392.5 ±29.9</td>
<td>128.1 ±25.4</td>
<td>409.1 ±161.8</td>
<td>2.5 ±1.07</td>
</tr>
<tr>
<td>ACH+DM</td>
<td>388.1 ±82.1</td>
<td>128.2 ±13.9</td>
<td>692.2 ±365.0***</td>
<td>3.2 ±2.50***</td>
</tr>
</tbody>
</table>

Control – rats receiving a normal diet and water; ACH – rats receiving low doses of ethanol; DM – streptozocin-treated rats; ACH+DM – rats exposed to low doses of ethanol and streptozocin; ALT – alanine aminotransferase; AST – aspartate aminotransferase; ALP – alkaline phosphatase; γ-GT – gamma-glutamyltransferase; the number of asterisks (*) or hashes (#) corresponds to the level of statistical significance (*** p < 0.001 combination vs control; # p < 0.05 combination vs ACH; ## p < 0.05 combination vs DM). All values refer to mean ± standard deviation (SD) serum levels, expressed in IU/L.

### Table 2. The lipid profile in diabetic and non-diabetic rats exposed to low-dose alcohol consumption

<table>
<thead>
<tr>
<th>Experimental group</th>
<th>TC</th>
<th>TG</th>
<th>HDL</th>
<th>LDL</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>51.1 ±15.2</td>
<td>131.2 ±58.9</td>
<td>34.1 ±3.4</td>
<td>17.3 ±10.5</td>
</tr>
<tr>
<td>ACH</td>
<td>48.6 ±13.1</td>
<td>132.1 ±71.3</td>
<td>31.1 ±2.8</td>
<td>20.7 ±6.6</td>
</tr>
<tr>
<td>DM</td>
<td>59.3 ±16.4</td>
<td>165.5 ±65.1</td>
<td>29.5 ±3.2</td>
<td>22.6 ±11.7</td>
</tr>
<tr>
<td>ACH+DM</td>
<td>80.5 ±24.6**</td>
<td>299.4 ±192.5**</td>
<td>24.5 ±12.6</td>
<td>38.8 ±16.3*</td>
</tr>
</tbody>
</table>

Control – rats receiving a normal diet and water; ACH – rats receiving low doses of ethanol; DM – streptozocin-treated rats; ACH+DM – rats exposed to low doses of ethanol and streptozocin; TC – total cholesterol; TG – triglyceride; HDL – high-density lipoprotein; LDL – low-density lipoprotein; the number of asterisks (*) or hashes (#) corresponds to the level of statistical significance (*** p < 0.001 combination vs control; # p < 0.05 combination vs ACH). All values refer to mean ± standard deviation (SD) serum levels, expressed in mmol/L.
a significant increase in all groups, with high levels in the combination group as an important pro-inflammatory mediator that acts as a protective marker in liver pathologies.\textsuperscript{13,25} On the other hand, some anti-inflammatory cytokines (IL-4 and IL-10) reacted positively. Interleukin-4 had an important role in protection against type 1 DM during pancreatic cell destruction and IL-10 level was non-significantly increased in response to Kupffer cell mediators with the inhibition of TNF-\(\alpha\).\textsuperscript{26,27}

The non-HDL lipid profile generally showed a significant increase by augmenting the effects of hypoinsulinemia and the lipolytic response with chronic low consumption of alcohol.\textsuperscript{28} High inflammatory cytokines also play an important role in lipid metabolism, stimulating the production of triglycerides in the liver.\textsuperscript{29–31} Hyperglycemia was still progressive, but to a lesser extent than in the DM group. This might be related to the hepatic insulin sensitizing effect of low doses of alcohol, which added to the low level of glucagon and decreased gluconeogenesis caused by alcohol in spite of the hyperglycemic effect of DM.\textsuperscript{17,32}

Therefore, a combined state of both low alcohol consumption and DM is associated with a depletion of the hyperglycemic response with elevated plasma cytokines, especially IL-6 and IL-4, which could be a part of a host defense mechanism to repair the hepatic and pancreatic damage through this inflammatory process. The severe liver damage under insult of low alcohol consumption and DM could serve as inhibitory factors in gluconeogenesis and glycogenolysis, with little or no impact on insulin levels.
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36. Dunning BE, Gerich JE. The role of alpha-cell dysregulation in fasting and postprandial hyperglycemia in type 2 diabetes and thera.