Do CTRC mutations affect the development of alcoholic chronic pancreatitis and its course among Poles: Preliminary study

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Abstract

Background. Genetic mutations are one of the etiological factors that predispose people to develop chronic pancreatitis.

Objectives. The aim of our study was to examine the effect of p.Trp55*, p.Arg254Trp and c.738_761del mutations in the chemotrypsin gene (CTRC) on the development of alcoholic chronic pancreatitis (ACP) in order to answer the questions whether these mutations vary between gender groups, whether they were related to the age when ACP was first diagnosed, and whether they affected the morphological changes in the pancreas and the course of ACP.

Material and methods. The study included 124 patients with ACP, 52 with nonalcoholic pancreatitis and 52 controls. The p.Trp55*, c.738_761del and p.Arg254Trp mutations in the CTRC gene were tested by the polymerase chain reaction (PCR).

Results. The c.738_761del and p.Arg254Trp mutations occurred in 3.07% and 1.31% of cases, respectively. None of the examined patients were found to have the p.Trp55* mutation. The frequency of detected mutations did not significantly differ between the study groups. The c.738_761del mutation was detected more frequently in women than in men. No significant differences were found in the age at ACP onset, morphological changes affecting the pancreas, or in the course of ACP between the patients with and without the 2 examined mutations. The c.738_761del mutation was significantly more frequent in the diabetic patients than in the non-diabetics. The patients with this mutation more frequently required surgery than those without the c.738_761del mutation.

Conclusions. No relationship between the c.738_761del and p.Arg254Trp mutations and the development of ACP was found. The c.738_761del mutation was more frequent in females than in males. Neither mutation affected the patient’s age at ACP onset or its course. In contrast to p.Arg254Trp, the c.738_761del mutation correlated with diabetes development and the need for surgery in the course of ACP.

Key words: alcohol, alcoholic chronic pancreatitis, CTRC mutations
Introduction

Chronic pancreatitis (CP) can present as recurrent episodes of acute inflammation or as progressive inflammatory conditions that result in fibrosis, calcification or altered morphology of the pancreas, the consequence of which is endocrine and exocrine failure of the organ. Global annual incidence rates of CP range from 5 to 12 cases per 100,000 people; the prevalence of CP is about 50 cases per 100,000 people. The disease occurs with a varied geographic distribution. In developed and western European countries, CP is generally due to alcohol consumption (38% of men and 11% of women). In the USA, Italy and Denmark, more than 50% of cases are alcohol-related, but in Brazil, the proportion reaches 89.6%. Alcohol consumption has been increasing in developing countries (China and India) due to rapid urbanization and increased affluence; therefore, this rise is expected to increase the burden of alcohol-related pancreatitis in these countries.

Among many etiological factors, the role of genetic predisposition to CP is clearly highlighted. The genetic factors of CP include a mutation/polymorphism of the cationic trypsinogen gene (PRSS1), the serine protease inhibitor Kazal type 1 gene (SPINK1), the chemotrypsin gene (CTRC), the cystic fibrosis transmembrane conductance regulator gene (CFTR), and the calcium sensing receptor gene (CASR). CASR is responsible for calcium homeostasis and some patients with hypercalcemia develop CP. This observation may encourage one to postulate that a mutation in the CASR gene may favor CP. Polymorphisms of interleukin-encoding genes, such as tumor necrosis factor-alpha (TNF-a), transforming growth factor-beta-1 (TGF-β1), interferon-gamma (INF-γ), vascular endothelial growth factor (VEGF), and intercellular adhesion molecule 1 (ICAM-1), are also major genetic contributors to the development of CP. In 2013, the carboxypeptidase A1 (CPA1) gene was identified as a novel gene of pancreatitis susceptibility. According to studies carried out on Polish individuals, the NS34 mutation of the SPINK1 gene seems to be significantly correlated with alcoholic chronic pancreatitis (ACP).

The CTRC gene encodes chemotrypsin C, a digestive enzyme produced by lobular pancreatic cells. Prematurely activated trypsin is destroyed by CTRC, which acts on the molecule within the calcium-binding loop in the absence of calcium, and therefore is a crucial candidate gene in the pathogenesis of CP. Since trypsin degradation serves as a protective mechanism against pancreatitis, it may be hypothesized that a loss of function in trypsin-degrading enzymes increases the risk for pancreatitis. Numerous CTRC mutations and polymorphisms have been presented so far. However, the p.Trp55*, p.Arg254Trp and c.738_761del mutations at exon 7 have not been studied among Poles with ACP, which poses a serious clinical challenge.

The aim of this study was to examine the effect that the p.Trp55*(W55X), p.Arg254Trp (R254W) and c.738_761del (del24) mutations in the CTRC gene have on the development of ACP, and to answer the question whether these mutations vary between gender groups. Moreover, we wanted to learn whether the mutations were related to the age when ACP was first diagnosed, and whether they affected the morphological changes in the pancreas and the course of ACP. The focus was on the 3 above-mentioned mutations of CTRC, because it seems that they have the greatest impact on the development of CP and are the most frequently observed mutations in exon 7.

Material and methods

Material

The study group was comprised of a homogenous Caucasian ethnic group of 228 patients, including 124 with ACP, 52 with nonalcoholic CP (NCP), and 52 healthy volunteers who made up the control group.

Chronic pancreatitis diagnosis was based on the standard criteria: a clinically confirmed history of recurrent episodes of acute pancreatitis and the results of imaging tests on the pancreatic structure (ultrasonography – USG, computed tomography – CT, endoscopy, or endoscopic retrograde cholangiopancreatography), such as calcifications, fibrosis of the pancreatic parenchyma, intraductal calcifications, and widened or irregular pancreatic ducts. Histological tests confirmed the diagnosis in 3 patients with ACP; in other cases, a biopsy was not performed. Alcoholic etiology was established on the basis of a medical history, i.e., the consumption of 80 g of pure ethanol in a 24-h period (males) or >40 g of pure ethanol in a 24-h period (females) in the previous 2 years or more. In addition, diabetes was diagnosed in 53 patients with ACP and insulin-dependent diabetes was diagnosed in 27 patients. Patients with a biliary, toxic, metabolic, or family history of CP were excluded from this group.

The group of patients with NCP included idiopathic CP cases. Chronic pancreatitis was diagnosed on the basis of the criteria presented above. The patients with an alcoholic, toxic, metabolic, or family history of CP were excluded from the study group. Four patients suffered from type 2 diabetes, treated with oral medication.

The control group was comprised of healthy volunteers with no history of alcohol consumption. They did not present any clinical symptoms or abnormalities on abdominal USG. There were no episodes of acute pancreatitis in this study group. Table 1 presents the characteristics of the study groups.

To examine whether there is an association between the p.Trp55*, p.Arg254Trp and c.738_761del mutations in the CTRC gene and the development of ACP, we compared the frequencies of the p.Trp55*, p.Arg254Trp and c.738_761del mutations in the CTRC gene in our 3 groups of patients: with ACP, with NCP, and in healthy controls.
The study protocol was approved by the local Ethical Committee (No. 131/2013) (Medical University of Lublin, Poland), and all participants gave written informed consent to participate in the study.

Methods

The p.Trp55*, c.738_761del and p.Arg254Trp mutations in the CTRC gene were tested in all patients.

DNA isolation

DNA was isolated from peripheral blood leukocytes, using a Blood DNA Purification Kit (EURx, Gdańsk, Poland) according to the instructions of the manufacturer. The lymphocytes were separated by the Ficoll gradient technique.

Determination of the p.Trp55* mutation in the CTRC gene

Polymerase chain reactions (PCRs) were performed with 100 ng of genomic DNA in a total volume of 20 μL, using a Biometra T Personal thermal cycler (Biometra, Gottingen, Germany). DNA was amplified with the primers named p.Trp55*_F and p.Trp55*_R (Table 2), designed by the Primer3 application in the Genetic Testing Laboratory in Lublin, Poland, and with a Taq PCR Master Mix kit (EURx), according to the manufacturers’ instructions. The setting parameters are listed in Table 2.

The PCR products were digested overnight at 37°C in a CLN 15 STD INOX/G incubator by the restriction enzyme PmlI (New England Biolabs, Ipswich, USA). The composition of the 20.4 μL restriction mix was 12 μL of the PCR product, 5 U of PmlI, 2 μL of CutSmart buffer, and 6 μL of water. Fifteen microliters of the digestion products were used for electrophoresis. The products of digestion were separated in 3% agarose gel (Sigma Aldrich, St. Louis, USA), stained with Simply Safe (EURx), and visualized on a transilluminator (JW Electronic, Warszawa, Poland). The wild alleles were digested into fragments of 202 bp and 262 bp; the mutated allele did not have a restriction site and was identified as a band of 462 bp.

To confirm the results, the analyzed samples underwent sequencing in both directions by means of primers 3R and 3F (Table 2), using a BigDye Terminator 3.1 kit (Life Technologies, Carlsbad, USA). The products of sequencing were electrophoretically separated on an ABI3100 sequencer, on a 50-centimeter capillary in the POP6 polymer, and analyzed by the Sequencing Analysis (Applied Biosystems, Foster City, USA).

Determination of the c.738_761del and p.Arg254Trp mutations in the CTRC gene

Both mutations were determined in 1 assay by using the same primer pairs, named c.738_761del_F and c.738_761del_R (Table 2), designed by Primer3 in the Genetic Testing Laboratory in Lublin, Poland. The PCR amplification was performed using a Biometra T Personal thermal cycler (Biometra) in a volume of 20 μL with 100 ng of genomic DNA, 2 primers and a Taq PCR Master Mix kit (EURx), according to the manufacturers’ instructions. The setting parameters are listed in Table 2. Following that, the PCR products were digested overnight at 37°C

Table 1. Characteristics of the study groups.

<table>
<thead>
<tr>
<th>Patient group</th>
<th>Number of patients</th>
<th>Gender</th>
<th>Age [years] mean±SD</th>
<th>Presence of diabetes [n]</th>
<th>Mean BMI [kg/m²] mean±SD</th>
<th>Mean age at CP onset [years] mean±SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>ACP</td>
<td>124</td>
<td>F 34</td>
<td>M 90</td>
<td>43.07 ±9.04</td>
<td>24.32 ±4.35</td>
<td>38.4 ±8.17</td>
</tr>
<tr>
<td>NCP</td>
<td>52</td>
<td>F 28</td>
<td>M 24</td>
<td>39.97 ±11.03</td>
<td>25.45 ±5.97</td>
<td>36.93 ±9.04</td>
</tr>
<tr>
<td>Controls</td>
<td>52</td>
<td>F 24</td>
<td>M 28</td>
<td>40.38 ±7.88</td>
<td>24.67 ±3.47</td>
<td>–</td>
</tr>
</tbody>
</table>

Table 2. Primers used to determine the p.Trp55*, p.Arg254Trp and c.738_761del mutations in the CTRC gene

<table>
<thead>
<tr>
<th>Application</th>
<th>Primer name</th>
<th>Primer sequence</th>
<th>Annealing temperature</th>
<th>Product length</th>
</tr>
</thead>
<tbody>
<tr>
<td>PCR-RFLP</td>
<td>p.Trp55*_F</td>
<td>5’ AGCCCTATTCCACTGGTTCTTCTTG 3’</td>
<td>59°C</td>
<td>464 bp</td>
</tr>
<tr>
<td></td>
<td>p.Trp55*_R</td>
<td>5’CAACTGAGTTACTGGGTGTGAGTAG 3’</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>c.738_761del_F</td>
<td>5’TGGTGCCCTATGCGCCCTCCG 3’</td>
<td>59°C</td>
<td>209 bp</td>
</tr>
<tr>
<td></td>
<td>c.738_761del_R</td>
<td>5’GGACAGCTGTGGAGGACGACGAC 3’</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sequencing</td>
<td>CTRC_3F</td>
<td>5’ACCTCGAGGCTGACACACA 3’</td>
<td>59°C</td>
<td>325 bp</td>
</tr>
<tr>
<td></td>
<td>CTRC_3R</td>
<td>5’GCTGGTTCTGGCAATACATAT 3’</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>CTRC_7F</td>
<td>5’CGAGCCAGGAGGCACAAAT 3’</td>
<td>58°C</td>
<td>467 bp</td>
</tr>
<tr>
<td></td>
<td>CTRC_7R</td>
<td>5’TGAATGAGTGGTGAATAGTGG 3’</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

PCR-RFLP – polymerase chain reaction-restriction fragments length polymorphism; both mutations, c.738_761del and p.Arg254Trp, were determined by using the same primers (c.738_761del_F and c.738_761del_R).
in a CLN 15 STD INOX/G incubator by the restriction enzyme SmaI (New England Biolabs). The composition of the 20.36 μL restriction mix was 12 μL of the PCR product, 10 U of SmaI, 1.6 μL of CutSmart buffer, 0.16 μL of bovine serum albumin (BSA), and 6 μL of water. Fifteen microliters of the digestion reaction products were used for electrophoresis. The products of digestion were separated in 3% agarose gel (Sigma Aldrich), stained with Simply Safe (EURx) and visualized on a transilluminator (JW Electronic). The wild allele was digested into fragments of 152 bp and 57 bp; the mutated allele (p.Arg254Trp) did not have restriction sites and appeared as a band of 209 bp. In the case of the c.738_761del mutation, the wild allele was seen as a band of 209 bp, but a product of 185 bp in length corresponded to the allele with a 24-bp deletion. To confirm the results, the analyzed samples underwent sequencing in both directions by means of primers 7F and 7R (Table 2) (the same primers for both mutations), using a BigDye Terminator 3.1 kit (Life Technologies) according to the manufacturer’s instructions. The products of sequencing were electrophoretically separated on an ABI3100 sequencer, on a 50-centimeter capillary in the POP6 polymer, and analyzed by the Sequencing Analysis (Applied Biosystems) according to the manufacturer’s instructions.

**Statistical analysis**

To describe the quantitative characteristics, the average values with standard deviations (SDs) were used. The comparison of the age at ACP onset was analyzed by Student’s t-test for independent samples. The qualitative data was described as numbers. To compare the frequency of the p.Trp55*, c.738_761del and p.Arg254Trp mutations in the CRTC gene between the study groups, the ANOVA test was performed. To compare the frequency of the examined mutations in the CRTC gene between women and men, the χ² test with Yates’s correction or Fisher’s exact test was used, as appropriate. Statistical significance between the differences was assumed at p < 0.05. All calculations were done by means of STATISTICA PL software (StatSoft, Kraków, Poland).

**Results**

The frequency of the examined mutations in the study groups is presented in Table 3. In the group of 228 patients, the c.738_761del mutation occurred in 3.07% of cases and the p.Arg254Trp mutation was found in 1.31% of patients (Fig. 1,2). In none of the examined patients was the p.Trp55* mutation detected. All mutations were heterozygotic. The c.738_761del mutation was detected only in the group of patients with ACP (4.35%); the p.Arg254Trp mutation was found in 1.45% of patients with ACP and in 1.92%

<table>
<thead>
<tr>
<th>Mutation</th>
<th>ACP</th>
<th>NCP</th>
<th>Controls</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>p.Trp55*</td>
<td>0/124</td>
<td>0/52</td>
<td>0/52</td>
<td>NS</td>
</tr>
<tr>
<td>c.738_761del</td>
<td>7/124</td>
<td>0/52</td>
<td>0/52</td>
<td>NS</td>
</tr>
<tr>
<td>p.Arg254Trp</td>
<td>2/124</td>
<td>0/52</td>
<td>1/52</td>
<td>NS</td>
</tr>
</tbody>
</table>

ACP – alcoholic chronic pancreatitis; NCP – nonalcoholic chronic pancreatitis; NS – no statistically significant differences.
of controls (Table 3). There were no statistically significant differences between the groups.

The c.738_761del mutation was detected in 6.98% of women and was statistically significantly more frequent in women than in men (0.70%) ($\chi^2 = 5.130; p = 0.0235$). The p.Arg254Trp mutation was detected in 1.16% of women and in 1.41% of men, and was not statistically significantly different ($\chi^2 = 0.025; p = 0.8746$).

The mean age at ACP onset in patients with the c.738_761del mutation was 39.87 ±5.56 years and without this mutation it was 38.22 ±8.15 years; with and without the p.Arg254Trp mutation, the mean age at ACP onset was 36.67 ±6.85 years and 38.94 ±7.87 years, respectively. The statistical analysis showed no statistically significant differences in the age at ACP onset between the patients with the c.738_761del and p.Arg254Trp mutations.

The relationship between the examined mutations and the course of ACP were analyzed with reference to morphological changes detected in the pancreas (calcifications, cysts, widening of the Wirsung duct by >3 mm, stones in the Wirsung duct), the occurrence of diabetes and the need for surgery. The patients with and without the c.738_761del and p.Arg254Trp mutations did not differ statistically in terms of morphological changes affecting the pancreas in the course of ACP. Fourteen patients were operated on: 2 had distal pancreatic resection, 7 had pancreaticoduodenectomy and 7 had cystojejunostomy. There were no statistical differences in the frequency of the examined mutations between patients operated on and not operated on in the course of ACP. However, the c.738_761del mutation was statistically significantly more frequent in patients with diabetes (13.21%) in the course of ACP compared to the examined non-diabetic patients (0%) ($p = 0.0204$); no differences were observed for the p.Arg254Trp mutation (Table 4).

## Discussion

Our results revealed a frequency of 5.65% and 2.42% for the c.738_761del and p.Arg254Trp mutations in the CTRC gene, respectively, in the group of patients with ACP compared to 0% and 1.92%, respectively, in the control group.

Rosendahl et al. studied the frequency of the del24 (c.738_761del) mutation in a group of German patients with ACP, and they found a 0.6% frequency of the del24 (c.738_761del) mutation in that group compared to 0.2% with alcoholic liver disease without pancreatitis. According to these authors, the frequency of the R2 54W (p.Arg254Trp) mutation in other types of CP was as follows: 1.7% in idiopathic CP, 4.2% in hereditary CP and 2.1% in healthy controls. However, the del24 (c.738_761del) mutation in the German patients occurred in 1.5% of idiopathic CP cases, and in 1.2% of controls; it was not detected in the hereditary CP group. Chemotrypsin C (CTRC) variants that diminish activity or secretion are associated with chronic pancreatitis. According to Felderbauer et al., the R2 54W (p.Arg254Trp) mutation occurred in 6.5% of the group of German patients with CP and hyperthyroidism, but was not detected in the group with hyperthyroidism without CP. The authors emphasized the correlation of that mutation type and the course of CP. However, they found no other mutation types in the CTRC gene. A study carried out in a group of patients with primary CP found a frequency of 1.7% for the R2 54W (p.Arg254Trp) mutation, 0.7% for the del24 (c.738_761del) mutation in a group of patients with idiopathic pancreatitis and 0.3% in healthy controls.

The research results of studies on CTRC mutations in CP etiology are controversial. Some researchers believe that CTRC mutations contribute to the so-called secondary CP (including this of alcoholic origin), while other genetic risk factors are either absent (e.g., PRSSI mutations) or they suggest a considerably lesser influence in comparison to primary CP (e.g., SPINK1 mutations). Therefore, those genes should be considered important contributing factors rather than causative factors. The studies in non-European populations found a higher frequency of the R2 54W (p.Arg254Trp) mutation in the group of patients with tropical pancreatitis (2.8%) compared to healthy volunteers (1.2%) of Indian origin. However, the del24 (c.738_761del) mutation, a considerably frequent type among German patients, was not found among Indian patients at all. Nonetheless, the interpretation of these results cannot be conclusive, because the Indian cohort was much smaller than the German group. The studies

### Table 4. Morphological changes in the pancreas and the clinical course of ACP in the patients with and without the c.738_761del and p.Arg254Trp mutations

<table>
<thead>
<tr>
<th>Course of ACP</th>
<th>c.738_761del</th>
<th>p.Arg254Trp</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>mutation (n)</td>
<td>no mutation (n)</td>
</tr>
<tr>
<td>Calcifications (n = 124)</td>
<td>7</td>
<td>117</td>
</tr>
<tr>
<td>Cysts (n = 54)</td>
<td>5</td>
<td>49</td>
</tr>
<tr>
<td>Widened Wirsung duct &gt;3 mm (n = 36)</td>
<td>4</td>
<td>32</td>
</tr>
<tr>
<td>Stones in Wirsung duct (n = 21)</td>
<td>3</td>
<td>18</td>
</tr>
<tr>
<td>Operation (n = 14)</td>
<td>5</td>
<td>9</td>
</tr>
<tr>
<td>Diabetes (n = 53)</td>
<td>6</td>
<td>47</td>
</tr>
</tbody>
</table>

ACP – alcoholic chronic pancreatitis; n – number of patients; NS – no statistically significant differences.
carried out in the Asian Pacific region found a significant correlation between the CTRC gene mutation and tropical calcific pancreatitis.\textsuperscript{18} Considering the biochemical activity of CTRC and the functional properties of the mutation, 3 mechanisms seem to contribute to the risk of developing CP: 1. a weakened degradation of trypsinogen and/or trypsin; 2. an impaired activation of A-carboxypeptidase; and 3. the induction of endoplasmic reticulum (ER) stress. Hence, the carriers of a CTRC-mutated gene are more likely to be exposed to ER stress in the exogenic region of the pancreas, which may contribute to apoptotic damage to the lobular pancreatic tissue.\textsuperscript{12,16,19}

The frequency of the c.738\_761del and p.Arg254Trp mutations in the CTRC gene among females and males has not been investigated so far. Our study seems to be the first in that respect. Our results found a statistically significantly higher frequency of the c.738\_761del mutation in females than in males. However, no such correlation was found for the p.Arg254Trp mutation.

Most research focuses on different roles of various mutations in the etiology of pancreatitis. When a mutation is confirmed, the course of CP is rarely examined in terms of environmental and demographic factors. It seems quite interesting to learn whether a CTRC mutation quickens and affects the course of CP, e.g., by developing diabetes. Our study found that a CTRC mutation did not affect the age at ACP onset or the course of the disease.

Moreover, diabetes correlated with the c.738\_761del mutation. There has been no research on CTRC gene mutations and CP course so far. However, the researchers have found that the N34S mutation in the SPINK1 gene favors ACP development and predisposes patients to developing diabetes at a younger age than patients without a mutated gene.\textsuperscript{5,11} We also found that patients with the c.738\_761del mutation more often required surgery in the course of ACP than patients without this mutation.

To our knowledge, this is the first study on the effect of the p.Trp55\*, p.Arg254Trp and c.738\_761del mutations in the CTRC gene on the development of ACP in Polish patients. Our results are preliminary and further large-population studies should be conducted to confirm these findings.

Conclusions

Our results lead to following conclusions:

1. No relationship between the c.738\_761del or p.Arg254Trp mutations in the CTRC gene and the development of APC or the course of APC was found in the examined group of Polish patients.

2. The mutation c.738\_761del occurred statistically significantly more frequently in women than in men; however, the p.Arg254Trp mutation was not gender-dependent.

3. Neither the c.738\_761del mutation nor the p.Arg254Trp mutation affected the patient’s age at ACP onset or the course of the disease.

4. Contrary to p.Arg254Trp, the c.738\_761del mutation in the CTRC gene correlated with diabetes development and with the need for surgery in the course of ACP.

References


