A large single-institution retrospective analysis of aggressive B-cell lymphomas according to the 2016/2017 WHO classification

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Abstract

Background. High-grade B-cell lymphomas (HGBLs) comprise a new entity in the revised 2016/2017 World Health Organization Classification of Tumours of Haematopoietic and Lymphoid Tissues. The diagnosis of HGBL encompasses histopathology and immunohistochemistry, with additional molecular examination of the BCL2/MYC or BCL6/MYC rearrangement status.

Objectives. The aim of the study was to summarize our experience in the histopathological and immunohistochemical diagnosis of patients with aggressive B-cell lymphomas according to the revised 2016/2017 WHO classifications.

Material and methods. We reviewed our single-institution experience with accurate diagnoses of HGBL and diffuse large B-cell lymphoma (DLBCL) using the available histopathological and immunohistochemical tools. The timeframe was from January 1, 2017 to April 18, 2018.

Results. Out of 265 patients, 217 (81.9%) were diagnosed with DLBCL, 43 (16.2%) with HGBL/DLBCL and 5 (1.9%) with not otherwise specified HGBL (HGBL-NOS). Regarding concurrent expression of MYC and BCL2 and/or BCL6 (double expressors (DE) and triple expressors (TE)), more DE and TE cases were found in the HGBL/DLBCL group than in the DLBCL group (25.53% vs 8.47%, p < 0.001, for DE cases and 55.32% vs 6.21%, p < 0.001, for TE cases). All 48 (100.00%) of the HGBL-NOS and HGBL/DLBCL patients, and 26 (11.98%) of the DLBCL-DE/TE cases were recommended for molecular analysis.

Conclusions. Our findings show that a comprehensive histopathological and immunohistochemical examination may identify potential HGBL cases. This study emphasizes the need to introduce a suitable molecular examination for patients with HGBL morphology and/or double/triple expression of BCL2/BCL6/MYC proteins.

Key words: DLBCL, World Health Organization (WHO) 2016/2017 Classification Of Tumours Of Haematopoietic and Lymphoid Tissues, DLBCL/BL, HGBL, high grade B-cell lymphoma/diffuse large B-cell lymphoma.

Received on August 27, 2018
Reviewed on March 10, 2019
Accepted on May 7, 2019
Published online on September 12, 2019
The revised 2016/2017 World Health Organization (WHO) Classification of Tumours of Haematopoietic and Lymphoid Tissues included new entities among aggressive B-cell lymphomas.\textsuperscript{1,2} The most significant changes were introduced by distinguishing a new category of high grade B-cell lymphomas (HGBLs), which were mainly derived from previous provisional categories, including unclassifiable B-cell lymphomas with features intermediate between diffuse large B-cell lymphoma (DLBCL) and Burkitt’s lymphoma (BL) (BCLU), as well as from DLBCLs of classic morphology.\textsuperscript{1,2}

The HGBL category was created based on the differences between its pathogenesis and clino-pathological features and those of DLBCL or BL.\textsuperscript{2–4} The poorer prognosis among HGBL patients in comparison to those with DLBCL or BL established a strong need to develop novel, more intensive therapies, as the standard regimen of rituximab, cyclophosphamide, vincristine, and doxorubicin (R-CHOP) is ineffective in HGBL patients.\textsuperscript{3,5–11} Although this new category is still highly heterogeneous, it creates an opportunity for a more accurate diagnosis and may ultimately lead to better treatment allocation or facilitate the search for new treatment strategies in randomized clinical trials.

An accurate diagnosis of HGBL encompasses standard histopathological and immunohistochemical analyses combined with molecular examination.\textsuperscript{2,4} Recently, Szumera-Cieckiewicz et al. comprehensively summarized the current practical guidelines on differential diagnosis of aggressive B-cell lymphomas, including the proper differentiation of HGBL.\textsuperscript{4} In brief, the current protocols for DLBCL/HGBL/BL diagnosis require the following to be determined: 1) the morphology of the lymphoma cells; 2) the cell of origin (COO), using the immunohistochemistry-based Hans protocol; 3) an immunohistochemical analysis of BCL2, BCL6 and MYC expression, as well as Tdt and cyclin D1/SOX11 in cases with blastoid morphology; and 4) a fluorescent in situ examination of BCL2/MYC or BCL6/MYC rearrangements (Table 1).\textsuperscript{1,2,4} While the first 3 requirements are widely available in Polish histopathology laboratories, there is a deficiency in access to proper molecular examinations.

In this study we aimed to conduct a large retrospective single-institutional report on the morphological and immunohistochemical diagnosis of aggressive B-cell lymphomas in light of the revised 2016/2017 World Health Organization (WHO) Classification of Tumours of Haematopoietic and Lymphoid Tissues.

The work described in this article was carried out in accordance with the Code of Ethics of the World Medical Association (Declaration of Helsinki) for experiments involving humans; EU Directive 2010/63/EU for animal experiments; and uniform requirements for manuscripts submitted to biomedical journals.

### Patients and methods

#### Study cohort

All aggressive B-cell lymphoma cases diagnosed at the Department of Pathology, Chair of Oncology, Medical University of Lodz, Poland, between January 1, 2017 and April 18, 2018 were included in the analysis. The review was restricted to cases classified as HGBL not otherwise specified (HGBL NOS – cases with clear blastoid morphology or not meeting the criteria for standard DLBCL or BL – Table 1), DLBCL and HGBL/DLBCL (a category for cases which in histopathological and immunohistochemical analysis appear to be HGBL, but no molecular confirmation was accessible). Formalin-fixed paraffin-embedded (FFPE) samples were collected, along with hematoxylin and eosin (H&E) and immunohistochemical slides. Finally, pathological data obtained from the patients was extracted into a pre-prepared Excel (v. 1907, Office 365; Microsoft Corp., Redmond, USA) spreadsheet. The required data encompassed the patient’s age at diagnosis, sex, diagnosis,

<table>
<thead>
<tr>
<th>Subtype</th>
<th>Morphology features</th>
<th>Immunohistochemistry</th>
<th>FISH analysis for BCL2/MYC or BCL6/MYC rearrangements</th>
<th>Most common diagnosis according to the previous 2008 WHO classification</th>
</tr>
</thead>
<tbody>
<tr>
<td>HGBL, NOS</td>
<td>Mandatory for diagnosis; blastoid or Burkitt-like (between BL and DLBCL)</td>
<td>GCB to ABC ratio ~ 2:1; DE/TE – most cases; negative for TdT and cyclin D1/SOX11</td>
<td>Not mandatory for diagnosis</td>
<td>DLBCL/BL</td>
</tr>
<tr>
<td>HGBL, DH/TH</td>
<td>Not mandatory for diagnosis; DLBCL – most cases, blastoid or Burkitt-like (between BL and DLBCL)</td>
<td>GCB – most cases; DE/TE – most cases; negative for TdT and cyclin D1/SOX11</td>
<td>Mandatory for diagnosis; positive for BCL2/MYC or BCL6/MYC rearrangement present</td>
<td>DLBCL</td>
</tr>
<tr>
<td>HGBL, DH/TH, transformed from FL</td>
<td>Mandatory for diagnosis; history of follicular lymphoma; DLBCL – most cases</td>
<td>GCB – most cases; DE/TE – most cases; negative for TdT and cyclin D1/SOX11</td>
<td>Mandatory for diagnosis; positive for BCL2/MYC or BCL6/MYC rearrangement present</td>
<td>DLBCL</td>
</tr>
</tbody>
</table>

NOS – not otherwise specified; DH – double-hit; TH – triple hit; GCB – germinal center B-cell phenotype; ABC – activated B-cell phenotype; DE – double expressor; TE – triple expressor. FISH – fluorescent in-situ hybridization; DLBCL/BL – unclassifiable B-cell lymphomas with features intermediate between diffuse large B-cell lymphoma (DLBCL) and Burkitt’s lymphoma (BL).
morphology, the results of immunohistochemical analysis including COO classification, Ki67 proliferation index, as well as BCL2, BCL6 and MYC expression.

Pathological diagnostic approach

Since January 1, 2017, our institution has followed a new diagnostic protocol for all aggressive B-cell lymphomas. The protocol is in accordance with the 2016/2017 WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues. In this protocol a diagnostic report of a lymphoma encompasses: 1) morphological texture (e.g., “high grade B-cell lymphoma with blastoid morphology”); 2) immunophenotype along with Ki67 proliferation index; 3) COO: germinal center B-cell phenotype (GCB) or activated B-cell phenotype (ABC), according to the Hans protocol\(^1\); 4) the status of immunoexpression of BCL2, BCL6 and/or MYC proteins (double expressor, triple expressor, non-double expressor); 5) a conclusion, including an indication whether the patient needs an additional molecular examination (e.g., “According to the 2016/2017 WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues, an additional fluorescence in situ hybridization (FISH) analysis should be performed on the sample for differential diagnosis between DLBCL and HGBL, DH/TH”). Examples of HGBL/DLBCL cases are shown in Fig. 1,2.

Immunohistochemistry

The standard panel of antibodies examined in patients with DLBCL, BL and HGBL covered CD20, CD3, BCL2, BCL6, MYC, CD10, MUM1, Ki67, cyclin D1, SOX11, TdT, CD5, CD38, and PAX5 (BSAP). Clones of the antibodies along with the manufacturers are listed in Table 2. Immunohistochemical analysis used monoclonal antibodies (FLEX Monoclonal Mouse Anti-Human, Dako A/S, Glostrup, Denmark) and EnVisionTM FLEX+ (Dako A/S) for the visualization. The tests were carried out using Autostainer Link 48 (Dako A/S).

The expression for all markers was reported as positive or negative, but for BCL2, BCL6, MYC, and Ki67 detailed

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**Fig. 1.** Histopathological example of a high grade B-cell lymphoma not otherwise specified (HGBL NOS) with the starry-sky pattern. The case was of germinal center B-cell phenotype origin and revealed BCL2 and MYC expression without expression of BCL6

DE – double expressor.
percentages of positive cells were reported in brackets. Cases were marked as positive under the following conditions: when more than 50% of the lymphoma cells were stained with anti-BCL2 antibody (cytoplasmic and nuclear staining), when more than 30% of the lymphoma cells were stained with BCL6 (nuclear staining) or when more than 40% of the lymphoma cells were stained with MYC (nuclear staining). The immunohistochemistry results were validated using positive and negative tissue controls in the whole series of immunostained slides.

**Statistical analysis**

Continuous variables were presented as medians followed by interquartile range (IQR), while nominal variables were presented as numbers followed by percentages in brackets. The Shapiro–Wilk test was used to assess the normality of distribution. Continuous variables were compared using the Mann–Whitney U test in case of non-normal distribution. Differences between categorical
variables were evaluated using the χ² or two-tailed Fisher’s exact tests. The STATISTICA v. 12.5 PL software package (Statsoft Inc., Tulsa, USA) was used for the analysis. P-values <0.05 were considered statistically significant.

Results

Patients characteristics

Between January 1, 2017 and April 18, 2018, 265 patients were diagnosed with either DLBCL (n = 217, 81.88%), HGBL, NOS (n = 5, 1.89%), or HGBL/DLBCL (n = 43, 16.23%). The median age of the whole group was 69 years (IQR = 61.12–77.77). The HGBL/DLBCL patients were older than DLBCL patients; however, this result was not statistically significant. Almost ½ of the patients in each group were males.

All HGBL NOS cases presented a blastoid-pattern morphology and were either double or triple expressors. Due to the small number of cases, all 5 HGBL NOS cases were included with the HGBL/DLBCL patients. The details of the study group, along with comparisons between the DLBCL and HGBL/DLBCL patients, are presented in Table 3.

<table>
<thead>
<tr>
<th>Variable</th>
<th>HGBL/DLBCL (n = 48, 18.12%)</th>
<th>DLBCL (n = 217, 81.88%)</th>
<th>p-value (test)</th>
<th>Whole group (n = 265, 100.00%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age [years]</td>
<td>72.16 (64.28–80.19)</td>
<td>68.73 (60.57–76.15)</td>
<td>0.086 (Mann–Whitney U test)</td>
<td>68.89 (61.12–77.77)</td>
</tr>
<tr>
<td>Sex (males)</td>
<td>21 (45.65%)</td>
<td>99 (46.26%)</td>
<td>0.940 (χ²)</td>
<td>120 (46.15%)</td>
</tr>
<tr>
<td>Hematology Center KMH EC</td>
<td>33 (68.75%)</td>
<td>99 (45.62%)</td>
<td>0.044 (χ²)</td>
<td>132 (49.81%)</td>
</tr>
<tr>
<td>HGBL, NOS</td>
<td>5 (10.41%)</td>
<td>NA</td>
<td>NA</td>
<td>5 (1.89%)</td>
</tr>
<tr>
<td>Morphology</td>
<td>6 (12.50%)</td>
<td>NA</td>
<td>NA</td>
<td>5 (2.26%)</td>
</tr>
<tr>
<td>Blastoid BL/DLBCL DLBCL</td>
<td>14 (29.17%)</td>
<td>245 (100.00%)</td>
<td></td>
<td>14 (5.28%)</td>
</tr>
<tr>
<td>Cell of origin GCB ABC</td>
<td>14 (29.17%)</td>
<td>82 (40.20%)</td>
<td>0.186 (χ²)</td>
<td>96 (37.23%)</td>
</tr>
<tr>
<td>BCL2 (positive)</td>
<td>41 (87.23%)</td>
<td>115 (59.90%)</td>
<td>&lt;0.001 (χ²)</td>
<td>156 (59.90%)</td>
</tr>
<tr>
<td>BCL6 (positive)</td>
<td>34 (72.34%)</td>
<td>137 (66.83%)</td>
<td>0.466 (χ²)</td>
<td>171 (67.89%)</td>
</tr>
<tr>
<td>MYC (positive)</td>
<td>38 (80.85%)</td>
<td>29 (16.48%)</td>
<td>&lt;0.001 (χ²)</td>
<td>67 (30.04%)</td>
</tr>
<tr>
<td>DE (positive) BCL2/MYC BCL6/MYC</td>
<td>9 (19.15%)</td>
<td>10 (5.65%)</td>
<td>0.005 (χ²)</td>
<td>19 (8.48%)</td>
</tr>
<tr>
<td>TE (positive)</td>
<td>26 (55.32%)</td>
<td>11 (6.21%)</td>
<td>&lt;0.001 (χ²)</td>
<td>37 (16.52%)</td>
</tr>
<tr>
<td>Ki67 index</td>
<td>95 (0.00–100.00)</td>
<td>90.00 (75.00–90.00)</td>
<td>&lt;0.001 (Mann–Whitney U test)</td>
<td>90.00 (80.00–95.00)</td>
</tr>
</tbody>
</table>

HGBL – high grade B-cell lymphoma; NOS – not otherwise specified; BL – Burkitt’s lymphoma; GCB – germinal center B-cell phenotype; ABC – activated B-cell phenotype; DLBCL – diffuse large B-cell lymphoma; NA – not applicable; KMH – Kopernik Memorial Hospital; EC – external oncological centers; DE – double expressor; TE – triple expressor.

Table 3. Characteristics of the HGBL/DLBCL and DLBCL study population. Quantitative variables are presented as medians followed by quartiles in brackets and as numbers followed by percentages in brackets.

Lymphoma morphology

Within the HGBL/DLBCL group, 6 patients (12.50%) presented with partially blastoid morphological features, and 14 patients (29.17%) presented with BL morphological features, while more than ½ presented with standard DLBCL morphology (mainly centroblastic or immunoblastic).

Comparison of cell of origin of lymphomas and Ki67 index

The majority of the lymphomas displayed an immunophenotype characteristic of ABCs. This tendency was stable regardless of the study group. There were 33 ABCs (70.21%) in the HGBL/DLBCL group and 122 ABCs (59.80%) in the DLBCL group (p = 0.186).

The Ki67 proliferation index in the whole study group was high: 90% (IQR = 80–95%). It differed significantly between HGBL/DLBCL and DLBCL patients: 95% (90–100%) vs 90.00% (75–90%), respectively (p < 0.001).

Comparison of BCL2, BCL6 and MYC expression

The majority of the lymphomas studied displayed positivity for BCL2 and BCL6 expression: n = 156 (65.27%) and n = 171 (67.86%), respectively; and 67 patients (30.04%) were positive for MYC expression. The percentage of BCL6-positive lymphomas did not differ significantly between the study groups (p = 0.466). However, HGBL/DLBCL cases were more frequently BCL2-positive and MYC-positive than DLBCL cases: 41 (87.23%) vs 115 (59.90%) BCL2-positive cases (p < 0.001) and 38 (80.85%) vs 29 (16.48%) MYC-positive cases (p < 0.001).

Regarding concurrent expression of MYC and BCL2 and/or BCL6, more DE and TE cases were found in the HGBL/DLBCL group than in the DLBCL group: 12 (25.53%) vs 15 (8.47%) DE cases (p < 0.001) and 26 (55.32%) vs 11 (6.21%) TE cases (p < 0.001); DE and TE cases comprised 54 (20.40%) of the patients in the study.

Patients recommended for molecular analysis

We recommended fluorescence in situ hybridization (FISH) analysis for identifying BCL2/MYC and
**Discussion**

During the 18 months following the introductions of the revised 2016/2017 WHO Classification of Tumors of Haematopoietic and Lymphoid Tissues, our clinic recommended almost 20% of HGBL patients and 30% of HGBL/DLBCL-DE patients for FISH analysis for proper discrimination between HGBL, DH/TH and DLBCL. Considering the important clinical and biological differences between these diagnoses, we demonstrated a pressing need to apply the differential diagnosis of DLBCL and HGBL in the routine diagnosis of lymphomas in Poland.

The HGBL category was formally introduced in 2017 with the publication of the new WHO classifications; however, this distinct subgroup of lymphomas had been proposed a few years earlier. The most important morphological criterion (the blastoid pattern) and the additional immunohistochemical criteria (double- or triple-expression of MYC, BCL2 and/or BCL6) are most often published in guidelines and reviews. Therefore, the present study separated all patients who were positive for the first or both criteria and labelled them as HGBL/DLBCL patients. No stringent criteria exist that can help to determine the additional molecular testing needed to separate HGBL DH/TH cases.

HGBL/DLBCL patients constituted almost 20% of our study group. This percentage is a little lower than literature values (in the largest cohorts, 30% of DLBCL cases were classified as HGBL/DLBCL; 6% were confirmed as HGBL-DH after molecular testing). In our study, the discrepancy was eliminated when DLBCL DE/TE cases were included in the group recommended for additional testing. The inclusion of DLBCL DE/TE for additional molecular assessment is clearly substantiated by recent reports on DLBCL DH cases within the DLBCL DE/TE group.

Most of the morphological and immunohistochemical characteristics identified in the group were consistent with those presented in previous reports; however, 2 were found to be intriguingly different. Firstly, almost 70% of the HGBL/DLBCL group displayed an ABC subtype phenotype when we assessed the COO using the Hans algorithm. The opposite GCB-to-ABC ratio is typically presented in the literature, with the value reaching as high as 100% GCB in true HGBL DH (BCL2/MYC rearranged) cases. The older age of the HGBL/DLBCL patients in our study (median age: 72 years) may explain this discrepancy, as ABC cases are more common among older patients. Another possible explanation is that our sample included a lower percentage of DE/TE cases among the ABC DLBCLs than reported previously.

Secondly, while a significantly higher Ki67 proliferation index was found in the HGBL/DLBCL group than in the DLBCL group in the present study, previous reports indicate no significant differences in this parameter, and advise against using it as a differential criterion for HGBL and DLBCL cases. We agree with these observations, because despite the statistical significance, the absolute differences in Ki67 proliferation index in our study were around 5%.

The need for detection of HGBL is reinforced not only by the different pathogenesis of HGBL and DLBCL, but also by the important clinical differences between HGBL and DLBCL mentioned previously. In comparison with DLBCL patients, HGBL (DH/TH or NOS) patients are characterized by shorter overall and event progression-free survival, and are more frequently associated with poor prognostic factors, such as age at diagnosis, high IPI scores and advanced disease. Moreover, diagnoses of HGBL (DH/TH or NOS) may soon become predictive for treatment allocation, especially among younger patients; most studies indicate that this group should be treated more intensively than with a standard R-CHOP regimen. However, it must be emphasized that trials of more intensive or novel regimens in HGBL have reported inconsistent results, and the evidence is still not strong enough to prepare distinct treatment guidelines for HGBL patients. This further emphasizes the need for better distinction of HGBL as a highly heterogeneous, provisional category of lymphomas, which should be investigated further.

The major limitation of our study is its lack of FISH analysis of HGBL/DLBCL and DLBCL DE/TE cases, which might be valuable for the final diagnosis. We plan to conduct this type of examination whenever possible in the future. A second limitation concerns the limited amount of clinical data and follow-up. We did not present these data in this report because the scope of the study was restricted to the diagnostic aspect of HGBL/DLBCL. In addition, our pathology department treats patients from several regional oncology centers and gathering such a large volume of data would be excessively time-consuming. Finally, the patients in our study were diagnosed following January 1, 2017 and no accurate conclusions could be drawn in such a short time since the diagnosis. We plan to update the report with clinical and molecular details in the near future.

**Conclusions**

Our findings show that comprehensive histopathological and immunohistochemical examinations can identify potential HGBL cases. As many as 20% of our HGBL/DLBCL patients would need FISH examination for BCL2/MYC or BCL6/MYC rearrangements. This is the strongest justification for the need to introduce appropriate examinations among patients with high grade B-cell lymphomas.
References


